Abstract

This document describes design decisions, and discusses implementation and algorithmic details in some vegan functions. The proper FAQ is another document.

Contents

1 Parallel processing 1
  1.1 User interface .................. 1  
    1.1.1 Using parallel processing as default .......... 1  
  1.1.2 Setting up socket clusters .. 2  
  1.1.3 Random number generation ................. 2  
  1.1.4 Does it pay off? ................. 2  
  1.2 Internals for developers ................. 2  

2 Nestedness and Null models 3  
  2.1 Matrix temperature ................. 3  

3 Scaling in redundancy analysis 4  

4 Weighted average and linear combination scores 5  
  4.1 LC Scores are Linear Combinations .......... 6  
  4.2 Factor constraints .................. 9  
  4.3 Conclusion .......................... 9  

1 Parallel processing

Several vegan functions can perform parallel processing using the standard R package parallel. The parallel package in R implements the functionality of earlier contributed packages multicore and snow. The multicore functionality forks the analysis to multiple cores, and snow functionality sets up a socket cluster of workers. The multicore functionality only works in unix-like systems (such as MacOS and Linux), but snow functionality works in all operating systems. Vegan can use either method, but defaults to multicore functionality when this is available, because its forked clusters are usually faster. This chapter describes both the user interface and internal implementation for the developers.

1.1 User interface

The functions that are capable of parallel processing have argument parallel. The normal default is parallel = 1 which means that no parallel processing is performed. It is possible to set parallel processing as the default in vegan (see §1.1.1). For parallel processing, the parallel argument can be either

1. An integer in which case the given number of parallel processes will be launched (value 1 launches non-parallel processing). In unix-like systems (e.g., MacOS, Linux) these will be forked multicore processes. In Windows socket clusters will be set up, initialized and closed.

2. A previously created socket cluster. This saves time as the cluster is not set up and closed in the function. If the argument is a socket cluster, it will also be used in unix-like systems. Setting up a socket cluster is discussed in §1.1.2.

1.1.1 Using parallel processing as default

If the user sets option mc.cores, its value will be used as the default value of the parallel argument in vegan functions. The following command will set up parallel processing to all subsequent vegan commands:

> options(mc.cores = 2)
The `mc.cores` option is defined in the `parallel` package, but it is usually unset in which case `vegan` will default to non-parallel computation. The `mc.cores` option can be set by the environmental variable `MC_CORES` when the `parallel` package is loaded.

R allows setting up a default socket cluster (`setDefaultCluster`), but this will not be used in `vegan`.

### 1.1.2 Setting up socket clusters

If socket clusters are used (and they are the only alternative in Windows), it is often wise to set up a cluster before calling parallelized code and give the pre-defined cluster as the value of the `parallel` argument in `vegan`. If you want to use socket clusters in unix-like systems (MacOS, Linux), this can be only done with pre-defined clusters.

If socket cluster is not set up in Windows, `vegan` will create and close the cluster within the function body. This involves following commands:

```r
clus <- makeCluster(4)
## perform parallel processing
stopCluster(clus)
```

The first command sets up the cluster, in this case with four cores, and the second command stops the cluster.

Most parallelized `vegan` functions work similarly in socket and fork clusters, but in `oecosimu` the parallel processing is used to evaluate user-defined functions, and their arguments and data must be made known to the socket cluster. For example, if you want to run in parallel the `meandist` function of the `oecosimu` example with a pre-defined socket cluster, you must use:

```r
> ## start up and define meandist()
> library(vegan)
> data(sipoo)
> meandist <-
> function(x) mean(vegdist(x, "bray"))
> library(parallel)
> clus <- makeCluster(4)
> clusterEvalQ(clus, library(vegan))
> sbcf <- oecosimu(dune, meandist, "r2dtable", parallel = clus)
> stopCluster(clus)
```

Socket clusters are used for parallel processing in Windows, but you do not need to pre-define the socket cluster in `oecosimu` if you only need `vegan` commands. However, if you need some other contributed packages, you must pre-define the socket cluster also in Windows with appropriate `clusterEvalQ` calls.

If you pre-set the cluster, you can also use `snow` style socket clusters in unix-like systems.

### 1.1.3 Random number generation

`Vegan` does not use parallel processing in random number generation, and you can set the seed for the standard random number generator. Setting the seed for the parallelized generator (L’Ecuyer) has no effect in `vegan`.

### 1.1.4 Does it pay off?

Parallelized processing has a considerable overhead, and the analysis is faster only if the non-parallel code is really slow (takes several seconds in wall clock time). The overhead is particularly large in socket clusters (in Windows). Creating a socket cluster and evaluating `library(vegan)` with `clusterEvalQ` can take two seconds or longer, and only pays off if the non-parallel analysis takes ten seconds or longer. Using pre-defined clusters will reduce the overhead. Fork clusters (in unix-like operating systems) have a smaller overhead and can be faster, but they also have an overhead.

Each parallel process needs memory, and for a large number of processes you need much memory. If the memory is exhausted, the parallel processes can stall and take much longer than non-parallel processes (minutes instead of seconds).

If the analysis is fast, and function runs in, say, less than five seconds, parallel processing is rarely useful. Parallel processing is useful only in slow analyses: large number of replications or simulations, slow evaluation of each simulation. The danger of memory exhaustion must always be remembered.

The benefits and potential problems of parallel processing depend on your particular system: it is best to rely on your own experience.

### 1.2 Internals for developers

The implementation of the parallel processing should accord with the description of the user interface above (§1.1). Function `oecosimu` can be used as a reference implementation, and similar
interpretation and order of interpretation of arguments should be followed. All future implementations should be consistent and all must be changed if the call heuristic changes.

The value of the parallel argument can be NULL, a positive integer or a socket cluster. Integer 1 means that no parallel processing is performed. The “normal” default is NULL which in the “normal” case is interpreted as 1. Here “normal” means that R is run with default settings without setting mc.cores or environmental variable MC_CORES.

Function oecosim interprets the parallel arguments in the following way:

1. NULL: The function is called with argument parallel = getOption("mc.cores"). The option mc.cores is normally unset and then the default is parallel = NULL.

2. Integer: An integer value is taken as the number of created parallel processes. In unix-like systems this is the number of forked multicore processes, and in Windows this is the number of workers in socket clusters. In Windows, the socket cluster is created, and if needed library(vegan) is evaluated in the cluster (this is not necessary if the function only uses internal functions), and the cluster is stopped after parallel processing.

3. Socket cluster: If a socket cluster is given, it will be used in all operating systems, and the cluster is not stopped within the function.

This gives the following precedence order for parallel processing (highest to lowest):

1. Explicitly given argument value of parallel will always be used.

2. If mc.cores is set, it will be used. In Windows this means creating and stopping socket clusters. Please note that the mc.cores is only set from the environmental variable MC_CORES when you load the parallel package, and it is always unset before first require(parallel).

3. The fall back behaviour is no parallel processing.

2 Nestedness and Null models

Some published indices of nestedness and null models of communities are only described in general terms, and they could be implemented in various ways. Here I discuss the implementation in vegan.

2.1 Matrix temperature

The matrix temperature is intuitively simple (Fig. 1), but the the exact calculations were not explained in the original publication (Atmar and Patterson 1993). The function can be implemented in many ways following the general principles. Rodríguez-Gironés and Santamaria (2006) have seen the original code and reveal more details of calculations, and their explanation is the basis of the implementation in vegan. However, there are still some open issues, and probably vegan func-
tion nestedtemp will never exactly reproduce results from other programs, although it is based on the same general principles. I try to give main computation details in this document — all details can be seen in the source code of nestedtemp.

- Species and sites are put into unit square (Rodríguez-Gironés and Santamaria 2006). The row and column coordinates will be \((k - 0.5)/n\) for \(k = 1 \ldots n\), so that there are no points in the corners or the margins of the unit square, and a diagonal line can be drawn through any point. I do not know how the rows and columns are converted to the unit square in other software, and this may be a considerable source of differences among implementations.

- Species and sites are ordered alternately using indices (Rodríguez-Gironés and Santamaria, 2006):
  \[
  s_j = \sum_{i|x_{ij}=1} i^2
  
  t_j = \sum_{i|x_{ij}=0} (n - i + 1)^2
  \]

  Here \(x\) is the data matrix, where 1 is presence, and 0 is absence, \(i\) and \(j\) are row and column indices, and \(n\) is the number of rows. The equations give the indices for columns, but the indices can be reversed for corresponding row indexing. Ordering by \(s\) packs presences to the top left corner, and ordering by \(t\) packs zeros away from the top left corner. The final sorting should be “a compromise” (Rodríguez-Gironés and Santamaria 2006) between these scores, and vegan uses \(s + t\). The result should be cool, but the packing does not try to minimize the temperature (Rodríguez-Gironés and Santamaria 2006). I do not know how the “compromise” is defined, and this can cause some differences to other implementations.

- The following function is used to define the fill line:
  \[
  y = (1 - (1 - x)^p)^{1/p}
  \]

This is similar to the equation suggested by Rodríguez-Gironés and Santamaria (2006, eq. 4), but omits all terms dependent on the numbers of species or sites, because I could not understand why they were needed. The differences are visible only in small data sets. The \(y\) and \(x\) are the coordinates in the unit square, and the parameter \(p\) is selected so that the curve covers the same area as is the proportion of presences (Fig. 1). The parameter \(p\) is found numerically using R functions integrate and uniroot. The fill line used in the original matrix temperature software (Atmar and Patterson, 1993) is supposed to be similar (Rodríguez-Gironés and Santamaria 2006). Small details in the fill line combined with differences in scores used in the unit square (especially in the corners) can cause large differences in the results.

- A line with slope \(-1\) is drawn through the point and the \(x\) coordinate of the intersection of this line and the fill line is found using function uniroot. The difference of this intersection and the row coordinate gives the argument \(d\) of matrix temperature (Fig. 1).

- In other software, “duplicated” species occurring on every site are removed, as well as empty sites and species after reordering (Rodríguez-Gironés and Santamaria, 2006). This is not done in vegan.

3 Scaling in redundancy analysis

This chapter discusses the scaling of scores (results) in redundancy analysis and principal component analysis performed by function rda in the vegan library.

Principal component analysis decomposes a centred data matrix \(X = \{x_{ij}\}\) into \(K\) orthogonal components so that \(x_{ij} = \sqrt{n - 1} \sum_{k=1}^K u_{ik} \sqrt{\lambda_k} v_{jk}\), where \(u_{ik}\) and \(v_{jk}\) are orthonormal coefficient matrices and \(\lambda_k\) are eigenvalues. In vegan the eigenvalues sum up to variance of the data, and therefore we need to multiply with the square root of degrees of freedom \(n - 1\). Orthonormality means that sums of squared columns is one and their cross-
product is zero, or \( \sum u_{ik}^2 = \sum v_{jk}^2 = 1 \), and
\[ \sum u_{ik}u_{il} = \sum v_{jk}v_{jl} = 0 \] for \( k \neq l \). This is a
decomposition, and the original matrix is found exactly
from the singular vectors and corresponding singular values, and first two singular components
give the rank = 2 least squares estimate of the original
matrix.

The coefficients \( u_{ik} \) and \( v_{jk} \) are scaled to unit
length for all axes \( k \). Eigenvalues \( \lambda_k \) give the
information of the importance of axes, or the ‘axis
lengths.’ Instead of the orthonormal coefficients, or
equal length axes, it is customary to scale species
(column) or site (row) scores or both by eigenval-
ues to display the importance of axes and to de-
scribe the true configuration of points. Table 1
shows some alternative scalings. These alterna-
tives apply to principal components analysis in all
cases, and in redundancy analysis, they apply to
species scores and constraints or linear combina-
tion scores; weighted averaging scores have some-
what wider dispersion.

In community ecology, it is common to plot both
species and sites in the same graph. If this graph
is a graphical display of PCA, or a graphical, low-
dimensional approximation of the data, the graph
is called a biplot. The graph is a biplot if the trans-
formed scores satisfy \( x_{ij} = c k(\sum w_{ij}^* v_{jk}^* ) \) where \( c \)
is a scaling constant. In functions princomp, pcomp
and rda with scaling = "sites", the plotted
scores define a biplot so that the eigenvalues are
expressed for sites, and species are left unscaled.

There is no natural way of scaling species and
site scores to each other. The eigenvalues in redu-
dancy and principal components analysis are scale-
dependent and change when the data are multiplied
by a constant. If we have percent cover data, the
eigenvalues are typically very high, and the scores
scaled by eigenvalues will have much wider disper-
sion than the orthonormal set. If we express the
percentages as proportions, and divide the matrix
by \( 100^2 \), the eigenvalues will be reduced by factor
100, and the scores scaled by eigenvalues will have
a narrower dispersion. For graphical biplots we
should be able to fix the relations of row and col-
umn scores to be invariant against scaling of data.
The solution in R standard function biplot is to
scale site and species scores independently, and typ-
ically very differently (Table 1), but plot each in-
dependently to fill the graph area. The solution in
Canoco and rda is to use proportional eigenval-
ues \( \lambda_k / \sum \lambda_k \) instead of original eigenvalues. These
proportions are invariant with scale changes, and
typically they have a nice range for plotting two
data sets in the same graph.

The vegan package uses a scaling constant \( c = \sqrt{(n-1)} \sum \lambda_k \) in order to be able to use scaling by
proportional eigenvalues (like in Canoco) and still
be able to have a biplot scaling. Because of this,
the scaling of rda scores is non-standard. However,
the scores function lets you to set the scaling con-
tant to any desired values. It is also possible to
have two separate scaling constants: the first for
the species, and the second for sites and friends,
and this allows getting scores of other software or
R functions (Table 2).

The scaling is controlled by three arguments in
the scores function in vegan:

1. scaling with options "sites", "species"
and "symmetric" defines the set of scores
which is scaled by eigenvalues (Table 1).

2. const can be used to set the numeric scaling
constant to non-default values (Table 2).

3. correlation can be used to modify species
scores so that they show the relative change
of species abundance, or their correlation with
the ordination (Table 1). This is no longer a
biplot scaling.

4 Weighted average and linear
combination scores

Constrained ordination methods such as Con-
strained Correspondence Analysis (CCA) and Re-
dundancy Analysis (RDA) produce two kind of site
scores [ter Braak 1986; Palmer 1993]:

- LC or Linear Combination Scores which are
  linear combinations of constraining variables.

- WA or Weighted Averages Scores which are
  such weighted averages of species scores that
  are as similar to LC scores as possible.

Many computer programs for constrained ordina-
tions give only or primarily LC scores following
recommendation of [Palmer 1993]. However, func-
tions cca and rda in the vegan package use primarily
WA scores. This chapter explains the reasons for
this choice.
Table 1: Alternative scalings for RDA used in the functions `prcomp` and `princomp`, and the one used in the `vegan` function `rda` and the proprietary software Canoco scores in terms of orthonormal species \((v_{ik})\) and site scores \((u_{ik})\), eigenvalues \((\lambda_k)\), number of sites \((n)\) and species standard deviations \((s_j)\). In `rda`, \(\text{const} = \sqrt{(n-1)\sum\lambda_k}\). Corresponding negative scaling in `vegan` is derived dividing each species by its standard deviation \(s_j\) (possibly with some additional constant multiplier).

<table>
<thead>
<tr>
<th>Function</th>
<th>Site scores (u_{ik}')</th>
<th>Species scores (v_{jk}')</th>
</tr>
</thead>
<tbody>
<tr>
<td><code>prcomp, princomp</code></td>
<td>(u_{ik}\sqrt{n-1}\sqrt{\lambda_k})</td>
<td>(v_{jk}\sqrt{n}\sqrt{\lambda_k})</td>
</tr>
<tr>
<td><code>stats::biplot</code></td>
<td>(u_{ik}\sqrt{n-1})</td>
<td>(v_{jk}\sqrt{\lambda_k})</td>
</tr>
<tr>
<td><code>stats::biplot, pc.biplot=TRUE</code></td>
<td>(u_{ik}\sqrt{\lambda_k}/\sum\lambda_k \times \text{const})</td>
<td>(v_{jk}\sqrt{\lambda_k}/\sum\lambda_k \times \text{const})</td>
</tr>
<tr>
<td><code>rda, scaling=&quot;sites&quot;</code></td>
<td>(u_{ik}\sqrt{\lambda_k}/\sum\lambda_k \times \text{const})</td>
<td>(v_{jk}\sqrt{\lambda_k}/\sum\lambda_k \times \text{const})</td>
</tr>
<tr>
<td><code>rda, scaling=&quot;species&quot;</code></td>
<td>(v_{jk}\sqrt{\lambda_k}/\sum\lambda_k \times \text{const})</td>
<td>(\sqrt{\sum\lambda_k/(n-1)s_j} v_{jk}')</td>
</tr>
<tr>
<td><code>rda, scaling=&quot;symmetric&quot;</code></td>
<td>(u_{ik}\sqrt{\lambda_k}/\sum\lambda_k \times \text{const})</td>
<td>(v_{jk}\sqrt{\lambda_k}/\sum\lambda_k \times \text{const})</td>
</tr>
<tr>
<td><code>rda, correlation=TRUE</code></td>
<td>(u_{ik}\times \text{const})</td>
<td>(v_{jk}\times \text{const})</td>
</tr>
</tbody>
</table>

Table 2: Values of the `const` argument in `vegan` to get the scores that are equal to those from other functions and software. Number of sites (rows) is \(n\), the number of species (columns) is \(m\), and the sum of all eigenvalues is \(\sum\lambda_k\) (this is saved as the item `tot.chi` in the `rda` result).

<table>
<thead>
<tr>
<th>Scaling</th>
<th>Species constant</th>
<th>Site constant</th>
</tr>
</thead>
<tbody>
<tr>
<td><code>vegan</code></td>
<td>any</td>
<td>(\sqrt{(n-1)\sum\lambda_k})</td>
</tr>
<tr>
<td><code>prcomp, princomp</code></td>
<td>1</td>
<td>(\frac{1}{\sqrt{n}})</td>
</tr>
<tr>
<td>Canoco v3</td>
<td>-1, -2, -3</td>
<td>(\frac{\sqrt{n-1}}{\sqrt{n}})</td>
</tr>
<tr>
<td>Canoco v4</td>
<td>-1, -2, -3</td>
<td>(\frac{\sqrt{n-1}}{\sqrt{m}})</td>
</tr>
</tbody>
</table>

Briefly, the main reasons are that

- LC scores are linear combinations, so they give us only the (scaled) environmental variables. This means that they are independent of vegetation and cannot be found from the species composition. Moreover, identical combinations of environmental variables give identical LC scores irrespective of vegetation.

- McCune (1997) has demonstrated that noisy environmental variables result in deteriorated LC scores whereas WA scores tolerate some errors in environmental variables. All environmental measurements contain some errors, and therefore it is safer to use WA scores.

This article studies mainly the first point. The users of `vegan` have a choice of either LC or WA (default) scores, but after reading this article, I believe that most of them do not want to use LC scores, because they are not what they were looking for in ordination.

4.1 LC Scores are Linear Combinations

Let us perform a simple CCA analysis using only two environmental variables so that we can see the constrained solution completely in two dimensions:

```r
> library(vegan)
> data(varespec)
> data(varechem)
> orig <- cca(varespec ~ Al + K, varechem)
> plot(orig, dis=c("lc","bp"))
```

Function `cca` in `vegan` uses WA scores as default. So we must specifically ask for LC scores (Fig. 2).

What would happen to linear combinations of LC scores if we shuffle the ordering of sites in species data? Function `sample()` below shuffles the indices.
It seems that site scores are fairly similar, but oriented differently (Fig. 3). We can use Procrustes rotation to see how similar the site scores indeed are (Fig. 4).

There is a small difference, but this will disappear if we use Redundancy Analysis (RDA) instead of CCA (Fig. 5). Here we use a new shuffling as well.

LC scores indeed are linear combinations of constraints (environmental variables) and independent of species data: You can shuffle your species data, or change the data completely, but the LC scores will be unchanged in RDA. In CCA the LC scores are weighted linear combinations with site totals of species data as weights. Shuffling species data in CCA changes the weights, and this can cause changes in LC scores. The magnitude of changes depends on the variability of site totals.

The original data and shuffled data differ in their goodness of fit:

```
> i <- sample(nrow(varespec))
> shuff <- cca(varespec[i,] ~ Al + K, varechem)
```

```
> plot(procrustes(scores(orig, dis="lc"),
               scores(shuff, dis="lc")))
```

```
> tmp1 <- rda(varespec ~ Al + K, varechem)
> i <- sample(nrow(varespec)) # Different shuffling
> tmp2 <- rda(varespec[i,] ~ Al + K, varechem)
```

```
```

The original data and shuffled data differ in their goodness of fit:

```
> orig
```
Figure 5: Procrustes rotation of LC scores in RDA of the original and shuffled data.

Call: cca(formula = varespec ~ Al + K, data = varechem)

<table>
<thead>
<tr>
<th>Inertia</th>
<th>Proportion</th>
<th>Rank</th>
</tr>
</thead>
<tbody>
<tr>
<td>Total</td>
<td>2.0832</td>
<td>1.000</td>
</tr>
<tr>
<td>Constrained</td>
<td>0.4760</td>
<td>0.2285</td>
</tr>
<tr>
<td>Unconstrained</td>
<td>1.6072</td>
<td>0.7715</td>
</tr>
</tbody>
</table>

Inertia is scaled Chi-square

Eigenvalues for constrained axes:

<table>
<thead>
<tr>
<th>CCA1</th>
<th>CCA2</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.3748</td>
<td>0.2404</td>
</tr>
<tr>
<td>0.1970</td>
<td>0.1782</td>
</tr>
<tr>
<td>0.1521</td>
<td>0.1184</td>
</tr>
<tr>
<td>0.0836</td>
<td>0.0757</td>
</tr>
</tbody>
</table>

(showing 8 of 21 unconstrained eigenvalues)

Similarly, their WA scores will be (probably) very different (Fig. 6).

The example used only two environmental variables so that we can easily plot all constrained axes. With a larger number of environmental variables the full configuration remains similarly unchanged, but its orientation may change, so that two-dimensional projections look different. In the full space, the differences should remain within numerical accuracy:

```r
> tmp1 <- rda(varespec ~ ., varechem)
> tmp2 <- rda(varespec[i,] ~ ., varechem)
> proc <- procrustes(scores(tmp1, dis="lc", choi=1:14), scores(tmp2, dis="lc", choi=1:14))
> max(residuals(proc))
[1] 3.356383e-14
```

In cca, the difference would be somewhat larger than now observed 3.3564e-14 because site weights used for environmental variables are shuffled with the species data.

Figure 6: Procrustes rotation of WA scores of CCA with the original and shuffled data.

Eigenvalues for unconstrained axes:

<table>
<thead>
<tr>
<th>CA1</th>
<th>CA2</th>
<th>CA3</th>
<th>CA4</th>
<th>CA5</th>
<th>CA6</th>
<th>CA7</th>
</tr>
</thead>
<tbody>
<tr>
<td>0.4740</td>
<td>0.3398</td>
<td>0.2101</td>
<td>0.1612</td>
<td>0.1343</td>
<td>0.1149</td>
<td>0.0924</td>
</tr>
<tr>
<td>CA8</td>
<td>0.0786</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

(showing 8 of 21 unconstrained eigenvalues)
4.2 Factor constraints

It seems that users often get confused when they perform constrained analysis using only one factor (class variable) as constraint. The following example uses the classical dune meadow data (Jongman et al., 1987):

\begin{verbatim}
> data(dune)
> data(dune.env)
> orig <- cca(dune ~ Moisture, dune.env)
\end{verbatim}

When the results are plotted using LC scores, sample plots fall only in four alternative positions (Fig. 7). In the previous chapter we saw that this happens because LC scores are the environmental variables, and they can be distinct only if the environmental variables are distinct. However, normally the user would like to see how well the environmental variables separate the vegetation, or inversely, how we could use the vegetation to discriminate the environmental conditions. For this purpose we should plot WA scores, or LC scores and WA scores together: The LC scores show where the site should be, the WA scores shows where the site is.

Function `ordispider` adds line segments to connect each WA score with the corresponding LC (Fig. 8).

\begin{verbatim}
> ordispider(orig, col="red")
> text(orig, dis="cn", col="blue")
\end{verbatim}

This is the standard way of displaying results of discriminant analysis, too. Moisture classes 1 and 2 seem to be overlapping, and cannot be completely separated by their vegetation. Other classes are more distinct, but there seems to be a clear arc effect or a “horseshoe” despite using CCA.

4.3 Conclusion

LC scores are only the (weighted and scaled) constraints and independent of vegetation. If you plot them, you plot only your environmental variables. WA scores are based on vegetation data but are constrained to be as similar to the LC scores as only possible. Therefore vegan calls LC scores as constraints and WA scores as site scores, and uses primarily WA scores in plotting. However, the user makes the ultimate choice, since both scores are available.
References


